



IMMUNOASSAYS AND SERVICES

BIOGENIC AMINES & NEUROSCIENCE | ENDOCRINOLOGY | FOOD SAFETY

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Instructions for use

Free Estriol ELISA

REF

FR E-2100



IVD



Free Estriol ELISA

1. INTENDED USE

The **Free Estriol ELISA** is an enzyme immunoassay for the quantitative *in vitro diagnostic* measurement of free estriol (unconjugated estriol) in serum during the second half of pregnancy.

This kit is NOT intended to be used for the risk evaluation of trisomy 21.

1.1 Summary and Explanation

Estriol (E3) is the major estrogen formed by the fetoplacental unit during pregnancy. Unconjugated E3 passes through the placenta into the maternal circulation, where it is rapidly converted into glucuronide and sulfate derivatives to facilitate its excretion. The half-life of estriol in the maternal bloodstream is only 20 – 30 minutes. Therefore, measurement of E3 offers a convenient and quick evaluation of current fetal status. Plasma estriol levels increase steadily throughout pregnancy and most rapidly during the third trimester (28 – 40 weeks). A sudden decrease in fetoplacental E3 production will result in a rapid decrease of unconjugated E3 in the maternal serum. There are several potential advantages to measuring unconjugated E3 rather than total serum or urinary E3. Unconjugated estriol levels are free from effects related to maternal renal or hepatic disease, and are not altered by the administration of certain antibiotics. Unconjugated E3 allows better prognosis in diabetic pregnancies - and since no hydrolysis of unconjugated E3 is required, a shorter time to first results.

2. PRINCIPLE OF THE TEST

The Free Estriol ELISA Kit is a solid phase enzyme-linked immunosorbent assay (ELISA), based on the **principle of competitive binding**.

The microtiter wells are coated with a polyclonal [rabbit] antibody directed towards an antigenic site on the Estriol molecule. Endogenous Estriol of a patient sample competes with an Estriol-horseradish peroxidase conjugate for binding to the coated antibody. After incubation the unbound conjugate is washed off.

The amount of bound peroxidase conjugate is inversely proportional to the concentration of Estriol in the sample. After addition of the substrate solution, the intensity of colour developed is inversely proportional to the concentration of Estriol in the patient sample.

3. WARNINGS AND PRECAUTIONS

1. This kit is for *in vitro* diagnostic use only. For professional use only.
2. All reagents of this test kit which contain human serum or plasma have been tested and confirmed negative for HIV I/II, HBsAg and HCV by FDA approved procedures. All reagents, however, should be treated as potential biohazards in use and for disposal.
3. Before starting the assay, read the instructions completely and carefully. Use the valid version of the package insert provided with the kit. Be sure that everything is understood.
4. The microplate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch and used in the frame provided.
5. Pipetting of samples and reagents must be done as quickly as possible and in the same sequence for each step.
6. Use reservoirs only for single reagents. This especially applies to the substrate reservoirs. Using a reservoir for dispensing a substrate solution that had previously been used for the conjugate solution may turn solution coloured. Do not pour reagents back into vials as reagent contamination may occur.
7. Mix the contents of the microplate wells thoroughly to ensure good test results. Do not reuse microwells.
8. Do not let wells dry during assay; add reagents immediately after completing the rinsing steps.
9. Allow the reagents to reach room temperature (21 °C to 26 °C) before starting the test. Temperature will affect the absorbance readings of the assay. However, values for the patient samples will not be affected.
10. Never pipet by mouth and avoid contact of reagents and specimens with skin and mucous membranes.
11. Do not smoke, eat, drink or apply cosmetics in areas where specimens or kit reagents are handled.
12. Wear disposable latex gloves when handling specimens and reagents. Microbial contamination of reagents or specimens may give false results.
13. Handling should be done in accordance with the procedures defined by an appropriate national biohazard safety guideline or regulation.
14. Do not use reagents beyond expiry date as shown on the kit labels.
15. All indicated volumes have to be performed according to the protocol. Optimal test results are only obtained when using calibrated pipettes and microtiter plate readers.
16. Do not mix or use components from kits with different lot numbers. It is advised not to exchange wells of different plates even of the same lot. The kits may have been shipped or stored under different conditions and the binding characteristics of the plates may result slightly different.









17. Avoid contact with *Stop Solution* containing 0.5 M H₂SO₄. It may cause skin irritation and burns.
18. Some reagents contain Proclin 300, BND and/or MIT as preservatives. In case of contact with eyes or skin, flush immediately with water.
19. TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them. If inhaled, take the person to open air.
20. Chemicals and prepared or used reagents have to be treated as hazardous waste according to the national biohazard safety guideline or regulation.
21. For information on hazardous substances included in the kit please refer to Safety Data Sheets. Safety Data Sheets for this product are available upon request directly from the manufacturer.

4. REAGENTS


4.1 Reagents provided


FR E-2131  96 **Microtiterwells**
 Content: 12 x 8 (break apart) strips; 96 wells; Wells coated with anti-Estriol antibody (polyclonal).

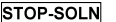
Standards and Controls - Ready to use

Cat. no.	Component	Standard	Concentration	Volume/Vial
FR E-2101	 STANDARD A	Standard A	0 ng/ml	1 ml
FR E-2102	 STANDARD B	Standard B	0.3 ng/ml	1 ml
FR E-2103	 STANDARD C	Standard C	1.2 ng/ml	1 ml
FR E-2104	 STANDARD D	Standard D	4.0 ng/ml	1 ml
FR E-2105	 STANDARD E	Standard E	15 ng/ml	1 ml
FR E-2106	 STANDARD F	Standard F	40 ng/ml	1 ml
FR E-2151	 CONTROL 1	Control 1	For control values and ranges please refer to vial label or QC-Report	1 ml
FR E-2152	 CONTROL 2	Control 2		1 ml

Contain non-mercury preservative.

FR E-2140  **Enzyme Conjugate** - Ready to use
 Content: Estriol conjugated to horseradish peroxidase; contains non-mercury preservative.
 Volume: 1 x 14 ml


ME E-0055  **Substrate Solution** - Ready to use
 Content: Tetramethylbenzidine (TMB)
 Volume: 1 x 14 ml

FR E-0080  **Stop Solution** - Ready to use
 Content: Contains 0.5 M H₂SO₄; avoid contact with the stop solution. It may cause skin irritations and burns.
 Volume: 1 x 14 ml

Hazards identification:



H290 May be corrosive to metals.
 H314 Causes severe skin burns and eye damage.

FR E-0030  **Wash Solution** - 40x concentrated
 Volume: 1 x 30 ml
 see "Preparation of Reagents".

Note: Additional Standard A for sample dilution is available upon request.

4.2 Materials required but not provided

- A calibrated microtiter plate reader (450 nm, with reference wavelength at 620 nm – 630 nm)
- Calibrated variable precision micropipettes
- Absorbent paper
- Distilled water
- Timer
- Semi logarithmic graph paper or software for data reduction

4.3 Storage Conditions

When stored at 2 °C to 8 °C unopened reagents will retain reactivity until expiration date. Do not use reagents beyond this date.

Opened reagents must be stored at 2 °C to 8 °C. Microtiter wells must be stored at 2 °C to 8 °C. Once the foil bag has been opened, care should be taken to close it tightly again.

Opened kits retain activity for 8 weeks if stored as described above.

4.4 Reagent Preparation

Bring all reagents and required number of strips to room temperature prior to use.

Wash Solution

Add distilled water to the 40X concentrated Wash Solution.

Dilute 30 ml of concentrated *Wash Solution* with 1170 ml distilled water to a final volume of 1200 ml.

The *diluted Wash Solution is stable for 1 week at room temperature.*

4.5 Disposal of the Kit

The disposal of the kit must be made according to the national regulations. Special information for this product is given in the Safety Data Sheet.

4.6 Damaged Test Kits

In case of any severe damage to the test kit or components, the manufacturer has to be informed in writing, at the latest, one week after receiving the kit. Severely damaged single components should not be used for a test run. They have to be stored until a final solution has been found. After this, they should be disposed according to the official regulations.

5. SPECIMEN COLLECTION AND PREPARATION

Serum should be used in this assay.

Note: Samples containing sodium azide should not be used in the assay.

In general it should be avoided to use haemolytic, icteric or lipaemic specimens. For further information refer to chapter "*Interfering Substances*".

5.1 Specimen Collection

Serum:

Collect blood by venipuncture (e.g. Sarstedt Monovette for serum), allow to clot, and separate serum by centrifugation at room temperature. Do not centrifuge before complete clotting has occurred. Patients receiving anticoagulant therapy may require increased clotting time.

5.2 Specimen Storage and Preparation

Specimens should be capped and may be stored for up to 7 days at 2 °C to 8 °C prior to assaying.

Specimens held for a longer time (up to 19 months) should be frozen only once at -20 °C prior to assay.

Thawed samples should be inverted several times prior to testing.

5.3 Specimen Dilution

If in an initial assay, a specimen is found to contain more than the highest standard, the specimens can be diluted with *Standard A* and reassayed as described in Assay Procedure.

For the calculation of the concentrations this dilution factor has to be taken into account.

Example:

a) dilution 1:10: 10 µl Serum + 90 µl *Standard A* (mix thoroughly)

b) dilution 1:100: 10 µl dilution a) 1:10 + 90 µl *Standard A* (mix thoroughly).

6. ASSAY PROCEDURE

6.1 General Remarks

- All reagents and specimens must be allowed to come to room temperature before use. All reagents must be mixed without foaming.
- Once the test has been started, all steps should be completed without interruption.
- Use new disposal plastic pipette tips for each standard, control or sample in order to avoid cross contamination.
- Optical density is a function of the incubation time and temperature. Before starting the assay, it is recommended that all reagents are ready, caps removed, all needed wells secured in holder, etc. This will ensure equal elapsed time for each pipetting step without interruption.
- As a general rule the enzymatic reaction is linearly proportional to time and temperature.

6.2 Test Procedure

Each run must include a standard curve.

1. Secure the desired number of Microtiter wells in the frame holder.
2. Dispense 10 µl of each Standard, Control and samples with <u>new disposable tips</u> into appropriate wells.
3. Dispense 100 µl Enzyme Conjugate into each well. Thoroughly mix for 10 seconds. It is important to have a complete mixing in this step.
4. Incubate for 60 minutes at room temperature.
5. Briskly shake out the contents of the wells. Rinse the wells 4 times with 400 µl diluted <i>Wash Solution</i> per well, if a plate washer is used - or - rinse the wells 4 times with 300 µl diluted <i>Wash Solution</i> per well for manual washing. Strike the wells sharply on absorbent paper to remove residual droplets. Important note: The sensitivity and precision of this assay is markedly influenced by the correct performance of the washing procedure!
6. Add 100 µl of Substrate Solution to each well.
7. Incubate for 30 minutes at room temperature.
8. Stop the enzymatic reaction by adding 100 µl of Stop Solution to each well.
9. Determine the optical density (OD) of the solution in each well at 450 nm (reading) and at 620 – 630 nm (background subtraction, recommended) with a microtiter plate reader. It is recommended that the wells be read within 10 minutes after adding the <i>Stop Solution</i> .

6.3 Calculation of Results

1. Calculate the average optical density (OD) values for each set of standards, controls and patient samples.
2. Using semi-logarithmic graph paper, construct a standard curve by plotting the mean OD obtained from each standard against its concentration with OD value on the vertical (Y) axis and concentration on the horizontal (X) axis.
3. Using the mean OD value for each sample determine the corresponding concentration from the standard curve.
4. Automated method: The results in the Instructions for Use have been calculated automatically using a 4 Parameter method curve fit. (4 Parameter Rodbard or 4 Parameter Marquardt are the preferred methods.) Other data reduction functions may give slightly different results.
5. The concentration of the samples can be read directly from this standard curve. Samples with concentrations higher than that of the highest standard have to be further diluted or reported as > 40 ng/ml. For the calculation of the concentrations this dilution factor has to be taken into account.

6.3.1 Example of Typical Standard Curve

The following data is for demonstration only and **cannot** be used in place of data generations at the time of assay.

Standard	Optical Density (450 nm)
Standard A (0.0 ng/ml)	1.79
Standard B (0.3 ng/ml)	1.48
Standard C (1.2 ng/ml)	1.18
Standard D (4.0 ng/ml)	0.81
Standard E (15.0 ng/ml)	0.52
Standard F (40.0 ng/ml)	0.38

7. EXPECTED NORMAL VALUES

It is strongly recommended that each laboratory should determine its own normal and abnormal values.

7.1 Normal healthy adults

In a study conducted with apparently normal healthy adults, using the Free Estriol ELISA the following values are observed:

Population	n	Mean [ng/ml]	Median [ng/ml]	5 th - 95 th Percentile [ng/ml]	2.5 th - 97.5 th Percentile [ng/ml]	Range (min. – max.) [ng/ml]
Males	42	0.359	0.367	0.146 - 0.573	0.077 - 0.878	0.075 - 0.987
Females (not pregnant)	43	0.348	0.286	0.057 - 0.822	0.038 - 1.027	0.009 - 1.307

The results alone should not be the only reason for any therapeutic consequences. The results should be correlated to other clinical observations and diagnostic tests.

7.2 Values during pregnancy

(p.m. = post menstruationem)

Week of gestation p.m.	Expected range [ng/ml]
12	0.3 - 1.0
13	0.3 - 1.1
14	0.4 - 1.6
15	1.0 - 4.4
16	1.4 - 6.5
17	1.5 - 6.6
18	1.6 - 8.5
19	1.9 - 11
20	2.1 - 13
21	2.6 - 14

Week of gestation p.m.	Expected range [ng/ml]	Twin pregnancy [ng/ml]
22 - 23	2.7 - 16	3 - 18
24 - 25	2.9 - 17	3 - 20
26 - 27	3.0 - 18	4 - 21
28 - 29	3.2 - 20	4 - 22
30 - 31	3.6 - 22	5 - 25
32 - 33	4.6 - 23	6 - 39
34 - 35	5.1 - 25	7 - 39
36 - 37	7.2 - 29	9 - 38
38 - 39	7.8 - 37	13 - 40
40 - 42	8.0 - 39	---

CLINICAL SIGNIFICANCE

The measurement of Estriol (E3) in body fluids has routinely been used for the monitoring and management of fetal well-being, particularly in the high-risk pregnant patient. The concentration of E3 in plasma increases gradually during the first 20 weeks gestation and more rapidly during the third trimester. Since the ranges for normal and abnormal serum conjugated E3 are wide and overlap considerably, a single E3 determination is of little value. The patient should be monitored frequently to establish any individual trend.

Consistently low levels or a sudden and continual decrease of serum E3 during the third trimester is highly indicative of fetal distress and possibly intrauterine death. When such observations are made, the status of the fetus should be assessed by alternative methods.

Interpretation of serum unconjugated E3 levels should be made in conjunction with other clinical tests or diagnostic procedures such as amniocentesis and ultrasound. Subnormal E3 levels may also be observed in patients being administered certain antibiotics or corticosteroids or in patients with impaired maternal hepatic function.

8. QUALITY CONTROL

Good laboratory practice requires that controls be run with each calibration curve. A statistically significant number of controls should be assayed to establish mean values and acceptable ranges to assure proper performance.

It is recommended to use control samples according to state and federal regulations. The use of control samples is advised to assure the day to day validity of results. Use controls at both normal and pathological levels.

The controls and the corresponding results of the QC-Laboratory are stated in the QC certificate added to the kit. The values and ranges stated on the QC sheet always refer to the current kit lot and should be used for direct comparison of the results.

It is also recommended to make use of national or international Quality Assessment programs in order to ensure the accuracy of the results.

Employ appropriate statistical methods for analysing control values and trends. If the results of the assay do not fit to the established acceptable ranges of control materials patient results should be considered invalid.

In this case, please check the following technical areas: Pipetting and timing devices; photometer, expiration dates of reagents, storage and incubation conditions, aspiration and washing methods.

After checking the above mentioned items without finding any error contact your distributor or the manufacturer directly.

9. PERFORMANCE CHARACTERISTICS

9.1 Assay Dynamic Range

The range of the assay is between 0.021 ng/ml – 40 ng/ml.

9.2 Specificity of Antibodies (Cross Reactivity)

The following substances were tested for cross reactivity of the assay:

Added steroid	Concentration of steroid	OD 450	Measured concentration
Estriol (E3)	40 ng/ml	0.39	39.67 ng/ml
Testosterone	16 ng/ml	1.758	n.d.
Estradiol (E2)	2 ng/ml	1.579	n.d.
Estrone (E1)	2 ng/ml	1.712	n.d.
Cortisol	800 ng/ml	1.775	n.d.

n.d. = non detectable

9.3 Sensitivity

The analytical sensitivity of the ELISA was calculated by subtracting 2 standard deviations from the mean of 20 replicate analyses of the Standard A and was found to be 0.021 ng/ml.

9.4 Reproducibility

9.4.1 Intra-Assay

The within-assay variability is shown below:

Sample	n	Mean [ng/ml]	CV (%)
1	20	2.1	4.7
2	20	6.2	3.2
3	20	14.6	3.0

9.4.2 Inter-Assay

The between-assay variability is shown below:

Sample	n	Mean [ng/ml]	CV (%)
1	12	2.1	4.6
2	12	5.7	8.5
3	12	13.3	9.5

9.5 Recovery

Recovery of the ELISA was determined by adding increasing amounts of the analyte to three different patient sera containing different amounts of endogenous analyte. Each sample (non-spiked and spiked) was assayed and analyte concentrations of the samples were calculated from the standard curve. The percentage recoveries were determined by comparing expected and measured values of the samples.

	Sample 1	Sample 2	Sample 3	
Concentration [ng/ml]	1.3	3.6	7.8	
Average Recovery	100.8	101.8	106.9	
Range of Recovery [%]	from	89.0	92.3	98.5
	to	103.8	109.8	112.3

9.6 Linearity

	Sample 1	Sample 2	Sample 3	
Concentration [ng/ml]	2.6	7.2	15.6	
Average Recovery	98.2	99.7	100.2	
Range of Recovery [%]	from	86.3	90.4	96.4
	to	107.9	106.8	103.7

10. LIMITATIONS OF USE

Reliable and reproducible results will be obtained when the assay procedure is performed with a complete understanding of the package insert instruction and with adherence to good laboratory practice. Any improper handling of samples or modification of this test might influence the results.

10.1 Interfering Substances

Haemoglobin (up to 4 mg/ml), bilirubin (up to 0.125 mg/ml) and triglyceride (up to 30 mg/ml) have no influence on the assay results.

10.2 Drug Interferences

Until today no substances (drugs) are known to us, which have an influence to the measurement of Estriol in a sample.

10.3 High-Dose-Hook Effect

No hook effect was observed in this test.

11. LEGAL ASPECTS

11.1 Reliability of Results

The test must be performed exactly as per the manufacturer's instructions for use. Moreover the user must strictly adhere to the rules of GLP (Good Laboratory Practice) or other applicable national standards and/or laws. This is especially relevant for the use of control reagents. It is important to always include, within the test procedure, a sufficient number of controls for validating the accuracy and precision of the test.

The test results are valid only if all controls are within the specified ranges and if all other test parameters are also within the given assay specifications. In case of any doubt or concern please contact the manufacturer.

11.2 Therapeutic Consequences

Therapeutic consequences should never be based on laboratory results alone even if all test results are in agreement with the items as stated under point 11.1. Any laboratory result is only a part of the total clinical picture of a patient.

Only in cases where the laboratory results are in acceptable agreement with the overall clinical picture of the patient should therapeutic consequences be derived.

The test result itself should never be the sole determinant for deriving any therapeutic consequences.

11.3 Liability












Any modification of the test kit and/or exchange or mixture of any components of different lots from one test kit to another could negatively affect the intended results and validity of the overall test. Such modification and/or exchanges invalidate any claim for replacement.

Claims submitted due to customer misinterpretation of laboratory results subject to point 11.2 are also invalid. Regardless, in the event of any claim, the manufacturer's liability is not to exceed the value of the test kit. Any damage caused to the test kit during transportation is not subject to the liability of the manufacturer.

12. REFERENCES/ LITERATURE

Bashore, R.A., Westlake, J.R. Plasma unconjugated estriol values in high risk pregnancy. Am. J. Obstet. Gynecol., June 15, 1977, p371-380

Symbols:

	Storage temperature		Manufacturer		Contains sufficient for <n> tests
	Expiry date		Batch code		For in-vitro diagnostic use only!
	Consult instructions for use		Content		CE labelled
	Caution		Catalogue number		